

Photo curiosity of the Portage Lake Association. Photographer, Peter Armstrong.

# **Portage Lake**

# Lake Management Plan 2017

Prepared for Onekama Township, Portage Lake Watershed Forever & Invasive Species Committee

Submitted By:

BreAnne Grabill, Environmental Scientist

# PLM Lake & Land Management Corp.

PO Box 424 · Evart, Michigan 49631 phone 800.382.4434 · fax 231.372.5900 www.plmcorp.net



# TABLE OF CONTENTS

Executive Summary	5
ntroduction	
Purpose of the Plan	5
Characteristics of the Lake	5
Management Goals for Portage Lake	6
Strategies for Achieving Lake Management Goals	
Aquatic Plant Control Techniques Chemical control Mechanical harvesting Biological control	8 8
BacteriaAeration	8
Integrated Pest Management (IPM)	
Exotic Plant Management	
Native Plant Management	
Algae Management	10
Monitoring	10
Nutrient Loading Abatement	10
Prevention	11
Lake Management Activities Conducted in 2017	11
Water Quality	11
Weather Challenges of 2017	
Aquatic Plant Control	
Map 1: Portage Lake June 2017 Treatment Map	12
Map 2: Portage Lake August 2017 Treatment Map	
Map 3: Portage Lake Terrestrial Treatment Map 2017  Table 1: Submersed Plant Treatment Quantities 2017-2009	13
Table 2: Terrestrial Treatment Summary (Phragmites, Narrow leaf cattails, Purple loos Japanese knotweed) 2017-2009	sestrife,
Planning/Evaluation Table 3: Plant Species Found in Portage Lake –2017	15
Graph 1: Native Plant Diversity (Fall AVAS Surveys)	16
Graph 2: EWM & Native Plant Cumulative Cover (C.C.) (Fall AVAS Surveys)	17
Genetic Testing/Sampling on Portage Lake	17
Current Conditions in the Lake	

Aquatic Vegetation	18
Water Quality Monitoring	19
Map 4: Portage Lake Water Quality Testing Locations	19
Table 4: Tributary Water Quality Portage Lake –2017 –cloudy/sprinkles/60	20
Table 5: Deep Hole Basin 1 Portage Lake –2017	20
(Secchi Disc: June 18', August 15', Sept.11')	
Table 6: Deep Hole Basin 2 Portage Lake -2017 (Secchi Disc: June 20', August 16', Sep	ot.
11')	21
Table 7: Shoreline Sampling Portage Lake –2017	22
Table 8: Additional shoreline sampling sites Portage Lake – 2017	22
Table 9: Storm Drain Sampling Portage Lake – May 1, 2017	22
Temperature and Dissolved Oxygen Profiles	
pH	24
Total Alkalinity	24
Conductivity and Total Dissolved Solids	24
Oxidative Reduction Potential (ORP)	24
Turbidity	2.5
Secchi Disk Depth	25
Graph 3: Spring Transparency (Secchi Disk) – Deep Hole Basins 1, 2 (1993-2017)	26
Graph 4: Fall Transparency (Secchi Disk) – Deep Hole Basins 1, 2 (1993-2017)	
Total Phosphorus	<u></u> 2 $\epsilon$
Graph 5: Total Phosphorus – Deep Hole Basins 1, 2 (2009-2017) (deep water sample)	27
Graph 6: Total Phosphorus & Dissolved Oxygen – Deep Hole Basin 1, (2009-2017) (dee	p
water sample)	28
Graph 7: Total Phosphorus & Dissolved Oxygen – Deep Hole Basin 2, (2009-2017) (dee	p
water sample)	28
Graph 8: Total Phosphorus – Tributaries 2009-2017	29
Graph 9: Total Phosphorus – Tributaries 2013-2017	29
Graph 10: Total Phosphorus – Tributaries May 2017	30
Graph 11: Total Phosphorus – Tributaries September 2017	
Graph 12: Total Phosphorus – Storm Drains May 2017	31
Graph 13: Total Phosphorus – Storm Drains May 2013 - 2017	31
Total Kjeldahl Nitrogen (TKN)	32
Graph 14: TKN – Portage Lake Basins 1, 2 (2009-2017) (deep water sample)	32
Graph 15: TKN & Dissolved Oxygen–Portage Lake Basin 1 (2009-2017) (deep water	
sample)	33
Graph 16: TKN & Dissolved Oxygen– Portage Lake Basin, 2 (2009-2017) (deep water	
sample)	33
Nitrates	33
Graph 17: Nitrates – Portage Lake Tributaries	34
Ammonia	34
Chlorophyll	
Graph 18: Chlorophyll a– Portage Lake Deep Basins	35
Algae and Zooplankton Composition	36
Fecal Indicator Bacteria (E. Coli)	36
Table 10: E. Coli Results in Portage Lake –2017	36
Tributary Flow and Phosphorus	37

Graph 19: Tributary Flow Rates –May and September 2013-2017	37
Graph 20: Tributary Flow Rates and Phosphorus (ug/L) comparisons –May 2017	38
Graph 21: Tributary Flow Rates and Phosphorus (ug/L) comparisons –September 2017 _	38
Additional Tributary/Upstream testing	38
Map 5: Portage Lake Upstream Tributary testing locations	39
Table 11: Upstream Tributary Testing 2017	
Graph 22: Total Phosphorus Stream #9	40
Evaluation of Trophic Status	40
Table 12: 2017 Trophic State Index (TSI) Values	
2017 Water Quality Concerns/Recommendations	41
Management Recommendations for 2018	41
Submersed Aquatic Plants	41
Emergent Vegetation Management	42
Monitoring	42
Proposed Budget	42
Table 13: Proposed 2018 Budget Portage Lake	42
The Recommended Management Schedule for 2018:	42

# Lake Management Plan

# **Executive Summary**

Portage Lake has been managed over the past nine years with goals of identifying and reducing the presence of exotic species throughout the Portage Lake watershed, tracking plant trends, improving water quality readings and protecting Portage Lake into the future. The following report breaks down the specifics of the previous management, the management of the 2017 season and the need for future management.

In 2017, just under 68 acres of EWM, Phragmites, Purple Loosestrife and Narrow leaf cattails were controlled via chemical control methods. Extensive lake mapping, vegetation mapping and water quality testing was also performed. The abundance of healthy native plants in Portage Lake increases the long term stability of the lake. While some water quality parameters have maintained themselves with little change over the years, other parameters have shown some fluctuations. One of the most important parameter to test is Total Phosphorus (TP). After finding a decrease in TP in 2015 and 2016, and a general downward trend, in 2017 TP concentrations increased. Although levels are only enriched and not highly enriched, the two-year trend down was not found again in 2017. Some of these fluctuations in other parameters include showing that the tributaries around Portage Lake are bringing excess nutrients into the lake. This information is vital in determining the areas within Portage Lake that need to be focused on reducing nutrient loading to help reduce the productivity in Portage Lake. The ability of Portage Lake to produce algae and aquatic plants is directly related to the overall health and use of Portage Lake. While the main goal of the management is to protect the long term ecological health of the lake, it is also important to protect the recreational, aesthetical and financial aspects of the lake as well. All of these factors play into the management efforts on Portage Lake, which need to be continued into next season.

Portage Lake was selected to be a sampling lake in PLM's DNR Grant study in 2015. PLM has collaborated with Michigan Tech University in a 3-year study to genetically test milfoil plants to determine the plant response to various chemical herbicides. This exciting study is still underway but should assist with management decisions and the direction of the program in the future. There is no current update available at the time the LMP was finalized.

#### Introduction

# **Purpose of the Plan**

This management plan documents management activities during 2017, examines current conditions in the lake, and provides management recommendations for 2018. The plan will detail an integrated approach to lake management including but not limited to exotic weed control, water quality monitoring and aquatic vegetation surveying.

#### Characteristics of the Lake

Portage Lake is a 2165-acre lake located in Onekama Township and the Village of Onekama, Manistee County, Michigan. Public access to the lake is provided by multiple access sties. A large portion of the shoreline has been developed and of that, a majority for single-family year-round homes. A formal lakeuse survey was not included in this study, but observations made while working on the lake indicate that the lake is used for fishing, boating (power & non-power), and swimming.

Portage Lake makes up 13.6% of the overall Portage Lake Watershed, which drains into Lake Michigan. Numerous other lakes tributaries flow into Portage Lake. which has a man-made channel into Lake Michigan on the west end. Portage Lake is a natural lake with two deep holes approximately 60' deep.

A few problems necessitating management of Portage Lake are:



(1) exotic and invasive species, and (2) water quality concerns. The presence of multiple exotic species has required annual management of the aquatic and terrestrial plants within and around Portage Lake.

Establishment of weedy exotic aquatic plants, including Eurasian watermilfoil and curly leaf pondweed, exacerbates problems caused by aquatic vegetation in the lake. These weedy exotic plants grow to the surface and cause substantially more interference with recreation than native plants.

# **Management Goals for Portage Lake**

- The primary goal of management in Portage Lake is to control and manage exotic plants, to allow
- recreational use of the lake and promote a healthy fishery. The exotic plant species, Eurasian watermilfoil and Phragmites, should be controlled throughout Portage Lake to the maximum extent possible. Native plants should be encouraged throughout the lake to promote an overall heahlty ecosystem. Genetic testing in Portage Lake has found that the Eurasian watermilfoil and Northern watermilfoil species have bred, forming a new genetic strand of milfoil commonly referred to as Hybrid milfoil. In reference to Portage Lake, Eurasian milfoil will be now referring to both EWM and Hyrbid milfoil as it all needs to be managed as an exotic, invasive species.



- Aquatic plant management should preserve species diversity and cover of native plants sufficient to provide habitat for fish and other aguatic organisms. Native plants should be managed to encourage the growth of plants that support the Portage Lake fishery (by creating structure and habitat) provided that they do not excessively interfere with recreational uses of the lake (e.g., swimming and fishing) in high-use areas. Where they must be managed, management techniques that reduce the stature of native plants without killing them (e.g.,
  - harvesting, contact herbicides) should be used whenever possible. Specific areas should be set aside where native plants will not be managed, to provide habitat for fish and other aquatic organisms. Muskgrass (Chara) should be allowed to grow throughout the lake, except in where it grows so tall as to interfere with boating and swimming.
- The species Starry stonewort, if found on the Portage Lake should be actively controlled and managed. Starry stonewort is in the same family as Muskgrass (Chara) but is considered an exotic invasive species. Starry stonewort, which looks very similar to the beneficial species Chara, is appearing in more and more lakes. Chara is a highly desired plant because it is typically low growing, keeps the water clear and can slow down the invasion of exotic weed species. Starry stonewort also forms dense mats, but



Starry stonewort

unlike chara, it can grow from 5 to 7 feet tall. Starry stonewort can be very detrimental to a lake's ecosystem and has the ability to kill off native plants and have a negative impact on a lake's fisheries.

The aquatic invasive terrestrial plants, Purple loosestrife and Phragmities should be controlled along



**Phragmites** 

the shoreline and adjacent wetlands where present. Both species are exotic and have the ability to displace beneficial native vegetation. Purple loosestrife grows 2 -4 feet tall and is a vibrant magenta color. It is very aggressive and can quickly become the dominant wetland vegetaion. Phragmites (common reed) is a wetland grass that ranges in height from 6 to 15 feet tall. "Phrag" quickly becomes the dominant feature in aquatic ecosystems, aggressively invading shorelines, wetlands, and ditches. This plant creates dense "strands" - walls of weeds crowding out beneficial native wetland vegetation and indigenous

waterfowl habitats. Spreading by fragmentation and an extensive root system, Phragmites ultimately out-competes native plant life for sun, water and nutrients.

The terrestrial invasive plant, Japanese knotweed should be controlled throughout the Portage Lake Watershed. Japanese knotweed is a large, herbaceous perennial plant native to Eastern Asia. In North America, the species has been classified as an invasive species. Japanese knotweed has hollow stems with distinct raised nodes that give it the appearance of bamboo, though it is not closely related. Reaching a maximum height of about 12' each growing season, it is typical to see much smaller plants in places where they sprout through cracks in the pavement or are repeatedly cut

down. The invasive root system and strong growth can damage concrete foundations, buildings, roads, paving, retaining walls and architectural sites. It can also reduce the capacity of channels to carry water. It forms thick, dense colonies that completely crowd out any other herbaceous species. The success of the species has been partially attributed to its tolerance of a very wide range of soil types, pH and salinity. The plant is also resilient to cutting, vigorously resprouting from the roots. The most effective method of control is by herbicide application close to the flowering stage in late summer or autumn.



Narrow-leaf cattails, another terrestrial invasive species, which can often be confused with the



Common cattail, are often found growing in marches, lakeshores, ponds, ditches, etc. Similar to other invasive species, Narrow-leaf cattails often form monocultures and outcompete other native species, leading to a concern for species habitat and often affecting recreational use of the area. Narrow-leaf cattail's leaves are about ½ inch wide, roughly half the width of the native broadleaf cattail. The stem is roughly 3-6' tall. The two species also hybridize, producing a cross that can exhibit characteristics of both species, though is often taller and more aggressive than either parent species and can be more difficult to identify. Management options include mowing, digging, grazing, water level manipulation, and chemical control.

- Water quality efforts in Portage Lake should continue to be made to reduce external loading of nutrients. Proper watershed management techniques should be applied where possible and lake residents should be encouraged to practice "lake friendly" lawn maintenance.
- Outreach/education of the Portage Lake residents should continue in an attempt to communicate lake activities and management goals. The Portage Lake website should be maintained as a way to directly relay pertinent information along with annual meetings and newsletters.

# **Strategies for Achieving Lake Management Goals**

### **Aquatic Plant Control Techniques**

Areas of the lake that support vegetation will grow plants, despite intense efforts to remove them. Aquatic vegetation provides important benefits to a lake, including stabilizing sediments, providing habitat for fish and other aquatic organisms, and slowing the spread of exotic plant species. In general, native plants interfere less with recreation and other human activities than exotic species. The nonnative plant species, Eurasian watermilfoil and curly leaf pondweed concentrate their biomass at the water surface where they strongly interfere with boating, swimming and other human activities. This growth form also allows exotic plants to displace native plants and form a monospecific (i.e., single species) plant community. The dense surface canopies of Eurasian watermilfoil and Curly leaf pondweed provide a lower quality habitat than that provided by a diverse community of native plants. Control of exotic plant species minimizes interference of plant growth with human activities and protects the native vegetation of the lake. The goal of environmentally responsible aquatic plant management, therefore, is not to remove all vegetation, but to control the types of plants that grow in the lake and the height of plants, to minimize interference with human activities.

It is important that control techniques meet the needs and expectations of lake users. Each technique has advantages and disadvantages. Many aquatic plants are relatively susceptible to some control measures but resistant to others. Too often, lake groups select a control technique before determining what their needs are.

Chemical control, or use of aquatic herbicides, is the most common strategy for controlling exotic plant species. Aquatic herbicides provide predictable results and there is a great deal of research and data regarding theses products. Many of the aquatic herbicides available can be used to selectively control exotic species with minimal or no impact on native species.

Mechanical harvesting is best suited for native plant species. Most native plant species have a higher

tolerance to aquatic herbicides and require higher dosage rates (higher cost and reduced selectivity). Mechanical harvesting can be used to provide relief from native plant species if they are causing a recreational nuisance. Harvesting does not kill the plants, but simply reduces it's stature, leaving lower growth for fish habitat and sedimnet stabilization. Mechanical harvesting of Eurasain watermilfoil is not recommended as it will expedite its spread throughout a lake through fragmentation.



Biological control options for nuisance aquatic vegetation are limited. Grass carp, which indiscriminately devour aquatic vegetation, have been restricted in many states because of their nonselective grazing and fear they may escape into nonintended waters. The use of the milfoil weevil (Euhrychipsis lecontei) to control Eurasian watermilfoil has been implemented in many Michigan lakes. PLM Lake & Land Management Corp has many years of experience particapating in weevil stocking, evaluations and longterm observations related to their performance and sustainability. Although the milfoil weevils may impact EWM populations in certain situations, the use of this tool remains unpredictable.

Bacteria product formulations and application techiques has greatly improved in recent years. Granular bacteria products can be applied to specific shoreline areas to reduce organic muck that has acumulated over the years. As waterbodies age, organic sediment can build up due to excessive plant and algae growth. This process is called eutrohpication. Increasing native populations of bacteria can slow this process down. Reductions in the depth of muck may depend on many variables. Most importantly, the

percent of sediment that is organic. The more organics in the sediment, the greater the potential for muck reduction via bacteria augmentation.

Aeration can be a beneficial tool to sustain ecological balance within an aquatic ecosystem. By

maintaining sufficient oxygen levels throughout a waterbody, the entire eutrophication process can be slowed down, the health of the fishery can be maintained and overall water quality can be improved. The implementation of an aeration system to control rooted aquatic plant growth is not recommended. Rooted plants, such as Eurasian watermilfoil, will not be affected by aeration. Similar to the use of biological control, the impact of aeration on improving water quality and reducing organic sediment will vary greatly from site to site. Therefore, it is extremely important to thoroughly evaluate each site's conditions and expectations before implementing an aeration system.



Integrated Pest Management (IPM) approaches to aquatic plant control IPM emphasize spending more effort evaluating the problem, so that exactly the right control can be applied at just the right time to control the pest. IPM approaches minimize treatment costs and the use of chemicals. Lake Management planning ensures the most appropriate, cost-effective treatment for your lake. Planning is an essential phase of Integrated Pest Management and includes lake vegetation surveys, water quality evaluation and a detailed, written lake management plan. Having the plan in place helps lake users know what to expect from lake management. Survey results provide a permanent record of conditions in the lake and the impact of management practices.

#### **Exotic Plant Management**

Aquatic herbicides currently represent the most reliable, effective, selective means for controlling Eurasian watermilfoil. There are currently five systemic herbicides, 2,4-D (Navigate), 2,4-D amine (Sculpin G), triclopyr (Renovate 3 & OTF), 2,4-D/Triclopyr combination (Renovate Max G) and fluridone (Sonar or Avast), which can be used to achieve long-term, selective control of Eurasian watermilfoil. Systemic herbicides are capable of killing the entire plant. Several contact herbicides, including diguat (Reward or Solera) can also provide short-term control of Eurasian watermilfoil. These herbicides kill only the shoots of the plant, and plants regrow relatively rapidly from their unaffected below ground parts.

Systemic herbicides control Eurasian watermilfoil with little or no impact on most native plant species. Under ideal conditions, several consecutive annual applications of these herbicides can reduce Eurasian watermilfoil to maintenance (low) abundance, such that only relatively small spot treatments are required to keep it under control. For this strategy to succeed, it is necessary to treat most of the Furasian watermilfoil in the lake each time.

Harvesting of Eurasian watermilfoil is not recommended. This plant spreads by fragmentation and regrows significantly more rapidly than most native plant species; thus continued harvesting of mixed plant beds typically leads to nearly complete domination of the aquatic vegetation by Eurasian watermilfoil.

Purple loosestrife can be selectively controlled through the use of triclopyr (Renovate). Purple loosestrife is an exotic species, which is out competing native vegetation, destroying valuable wetlands and animal habitat and expanding in density along Portage Lake. In past years our options to manage this nuisance weed has been extremely limited to prevention, manual removal or broad spectrum herbicide treatments, which not only killed the Purple Loosestrife but also the native vegetation remaining in the treatment areas. The biological control effort, beetles, have shown positive control measures and this method is also encouraged to continue into the future.

Phragmites, can be selectively controlled through the use of glyphosate or imazapyr (Habitat) herbicides. Phragmites is an exotic species, which can out compete native vegetation, destroying valuable wetlands and animal habitat.

# **Native Plant Management**

Native plants should be controlled primarily by harvesting if required. Unlike Eurasian watermilfoil, most native plants do not regrow rapidly after harvesting, and a single harvest is often sufficient to control them for the entire summer. Normally low-growing species should not be controlled unless unusually fertile growing conditions allow them to grow tall in areas of high recreational use. Contact herbicides applied at higher rates can be effective at controlling native plants that are causing a nuisance close to shore, in between docks.

# **Algae Management**

Not required at this time.

# **Monitoring**

It is important to maintain a record of lake conditions and management activities. Vegetation surveys monitor types and locations of plants in the lake, providing information that is essential to the administration of efficient, cost-effective control measures. Vegetation surveys also document the success or failure of management actions and the amount of native vegetation being maintained in the lake. Water quality monitoring can identify trends in water quality before conditions deteriorate to the point where remediation is prohibitively expensive or impossible. Records of past conditions and management activities also help to keep management consistent despite changes in the membership of the organization. Records should include (at a minimum):

- Temperature, dissolved oxygen and Secchi disk depth should be measured in the lake at both deep hole basins. Temperature and dissolved oxygen profiles should be obtained in the deep hole, so as to monitor the timing and extent of oxygen depletion in the hypolimnion (i.e., bottom water).
- Total phosphorus, nitrates, and ammonia should be measured in the surface and bottom water at least two times per season (spring and late summer) to monitor nutrient accumulation in the hypolimnion.
- Chlorophyll a sampling
- Tributary testing including flow and nutrient sampling
- Lake vegetation should be surveyed on an annual basis (late spring and/or late summer/early fall) to document the results of plant management efforts and provide information necessary for planning future management.

# **Nutrient Loading Abatement**

Lakeshore property owners should be encouraged to use phosphorus-free fertilizers on lawns and other areas that drain into Portage Lake or the adjacent wetlands. Lakeshore residents should also be encouraged to manage their waterside landscapes according to the recommendations outlined in publications on this topic available from the MSU Extension.

It is also important to remember that rooted plants derive most of their key nutrients from the sediments; thus, they respond slowly, if at all, to reductions in nutrient loading. In fact, if reductions in nutrient loading lead to improved water clarity, the growth of rooted plants will probably increase.

If organic material (muck) accumulates to undesirable levels in shoreline areas, bacterial treatments should be considered as a way to alleviate the buildup. PLM MD (Muck Digestion) Pellets are a combination of natural beneficial bacteria, enzymes, and vitamins that stimulate the biological activity of the lake bottom. This stimulation allows the bacteria to feed on the organic sediment, therefore reducing the muck levels over time.

#### **Prevention**

Eurasian watermilfoil and curly leaf pondweed were possibly introduced to Portage Lake by plant fragments carried on boats and/or boat trailers. A variety of other troublesome exotic plants and animals that have been introduced to Portage Lake are also transported this way. Preventing their inadvertent introduction to Portage Lake can significantly lower the cost of future lake management. Education can be an effective preventative measure. Newsletter articles should alert lake residents to the threat from exotic nuisance plants and animals. Warning signs should be erected at any public boat access sites, if applicable, that encourage boaters to clean boats and trailers when launching or removing watercraft from the lake.



# Lake Management Activities Conducted in 2017

#### **Water Quality**

Water quality was evaluated on May 1, June 1, August 2, and September 18, 2017. The May sampling included Storm Drain and tributary testing. In June, deep hole testing and shoreline testing of Portage Lake occurred. The August testing was for Ecoli, deep hole testing occurred (this was an additional sampling added into the program in 2015) and shoreline testing (added in 2017). During September, tributaries, shoreline and the deep hole basins were sampled. During the deep hole sampling the following occurred, (1) a depth profile of water temperature and dissolved oxygen concentrations were measured at ten feet intervals at both Deep Hole Basins and the Secchi disk depth was measured, (2) samples for LakeCheck™ analysis were collected from the deep holes of the lake (surface, bottom and every 10' between) for numerous parameters, (3) chlorophyll and algal composition analysis was collected from surface, mid thermocline and bottom samples. During the shoreline sampling, the following occurred, (1) depth profile for water temperature and dissolved oxygen concentrations were measured at the surface, (2) samples for LakeCheck™ analysis were collected at the surface for numerous parameters, (3) chlorophyll and algae composition analysis was collected at the surface. During the Storm Drain sampling the following occurred at four designated drains, (1) Flow testing, (2) surface reading of temperature and dissolved oxygen (3) samples for LakeCheck™ analysis were collected. During the tributary testing, the following occurred at seven designated tributaries, (1) surface reading for temperature and dissolved oxygen, (2) samples for LakeCheck™ analysis were collected and (3) flow was determined. LakeCheck measures at the various sites included some or all of the following parameters: Conductivity, Total Dissolved Solids, pH, Conductivity, Total Phosphorus, Oxidative Reduction Potential (ORP), Alkalinity, Ammonia, Nitrates and Total Kjeldahl Nitrogen. The additional tributary testing included sampling at one tributary and including testing multiple locations from the entrance at the lake, upstream. Parameters tested included Total Phosphorus, Nitrates and Alkalinity.

# Weather Challenges of 2017

Unlike many winters, which bring extremely cold temperatures with an above average snowfall, the 2016/2017 winter was similar to 2015/2016 with above average temperatures. Two above average winters will have short and long term impacts on the waterbody. Snow was late to come; lakes froze over later than normal and ice left in a hurry. Without a deep, hard freeze, many lakes do not slow production the same way they would on a cold winter. The summer brought warm temperatures, great for residents enjoying the water, but bringing difficulties to plant management programs. As the summer continued, the weather cooled in mid-late summer and August was one of the coldest on record. Despite these cooler temperatures, plants grew. Each lake responds differently from the weather impacts. Each year the weather will cause changes



Eurasian watermilfoil

within Portage Lake. Some years it may lower plant production while other years may lead to increased plant growth and elevated water quality numbers. Exotic species tend to benefit from changes in weather conditions. In Portage Lake, little plant growth was evident early on into the growing season and it was not until mid-summer that diverse plant coverage was found. Weather patterns can have impacts on lakes and individual plant trends that may not be evident right way, but months or years later. Although the mid/late summer temperatures were cool for recreational use, many plant species in Portage Lake thrived at those growing temperatures. The fall weather improved, with a warm September and plants continued to thrive until late into the year, which often leads to increased growth of exotic plants.

# **Aquatic Plant Control**

Weed treatments were conducted in June and August to control Eurasian watermilfoil (EWM) in Portage Lake. Phragmites and Narrow leaf cattails were also treated throughout 2017 around Portage Lake. The lake was closely monitored this year for any areas of exotic plant growth and treated accordingly.

The management strategy for the control of Eurasian watermilfoil has been working, with fewer acres of milfoil treated in 2017 than in 2015, 2014, 2013 or 2012. Although an



increase in acreage was found from the 2016 season, the overall level of milfoil is low. However, despite our efforts, EWM control is a constant battle that is heightened with hybrid watermilfoil. The presence of Hybrid watermilfoil supports the conclusion that milfoil treatments will continue to be required annually. A reflection of proper/successful management is a good fishery, which has been verified through the terrific fishing reports on the lake. Although fewer acres of milfoil treatment were required, the recommended application rates have increased, which uses up the budget more quickly. The Phragmites Treatment Program has been very effective. After the initial treatment of 83 acres, the follow up years have required just a small treatment in proportion to the initial application. The below maps and table show a breakdown of the treatments in Portage Lake in 2017, as well as previous years for both EWM and Phragmites/Purple Loosestrife/Japanese knotweed Control.





June 14, 2017 EWM and CLP Treatment, 1.5 acres Clipper at 200ppb

Map 2: Portage Lake August 2017 Treatment Map



August 15, 2017 EWM Treatment, 27 acres Renovate OTF (at 200 and 240lbs/acre) (purple on map), 33.5 acres Sculpin G (at 240lbs/acre) (yellow on map), 5.6 acres Renovate 3 (at 4 gals/acre) (red on map).

Map 3: Portage Lake Terrestrial Treatment Map 2017



October 6, 2017 Phragmites Treatment 0.15 acres within this 23.01 acre area

**Table 1: Submersed Plant Treatment Quantities 2017-2009** 

		Product	Rate#/Acre	Acres	Total Acres
2017	14- Jun	Clipper	200ppb	1.58	67.68
	15- Aug	Renovate OTF	200#	14	
			240#	13	
		Renovate 3	4gals	5.6	
		Sculpin G	200#	4	
		Sculpin G	240#	29.5	
2016	27-Jun	Clipper	200ppb	1.25	21.35
	2-Aug	Renovate OTF	200#	6.6	
		Renovate OTF	240#	3.5	
	3-Aug	Renovate OTF	200#	3	
		Renovate 3	4gals	2	
		Sculpin G	240#	5	
2015	6-Jun	Clipper	200ppb	1.25	79.35
	28-Jul	Renovate OTF	200#	4	
		Renovate OTF	240#	3.8	
		Sculpin G	200#	4	
		Sculpin G	240#	66.3	
2014	6-Jun	Renovate OTF	200#	1.5	176.05*
	29-Jul	Renovate OTF	200	.8	
		Renovate Max LZR	120#	95	
		Sculpin G	200#	10	
		Clipper	200ppb	1.25	
	8-Sep	Sculpin G	160#	23	
		Sculpin G	200#	12.5	
		Sculpin G	240#	6	
		Renovate Max LZR	160#	26	
2013	24,27 -Jun	Renovate OTF	160#	5	129.75
		Renovate Max G	160#	39	
		Sculpin G	160#	74.5	
	8-Aug	Sculpin G	160#	10	
		Clipper	200ppb	1.25	
2012	9-Jul	Renovate OTF	120#	10	145
		Renovate Max G	160#	55	
	24-Jul	Renovate OTF	120#	5	
		Renovate Max G	120#	40	
		Sculpin G (2,4-D)	160#	35	
2011	27-Jul	Renovate OTF	120#	22	22
2010	29-Jun	Renovate OTF	120#	5	86
		Navigate 2,4-D	100#	17	
	27-Sep	Renovate OTF	120#	14	
		Navigate 2,4-D	120#	50	
2009	15-Sep	Renovate OTF	120#	~41.5	161.5

Navigate 2,4-D	100#	120	
----------------	------	-----	--

\*Some Re-Treatment in 2014 due to in-adequate dieback of treatment beds.

Table 2: Terrestrial Treatment Summary (Phragmites, Narrow leaf cattails, Purple loosestrife, Japanese knotweed) 2017-2009

Year	Product	Rate	Acres
2017	Glyphosate/Imazapyr	1-3%	0.15
2016	Glyphosate/Imazapyr	1-3%	0.48
2015	Glyphosate/Imazapyr Triclopyr	1-3%	4
2014	Glyphosate/Imazapyr	4%	6.2
2013	Glyphosate/Imazapyr	2%	7.9
2012	Glyphosate/Imazapyr	2%	13.5
2011	Glyphosate/Imazapyr	2%	7
2010	Glyphosate/Imazapyr	2%	10
2009	Glyphosate/Imazapyr	2%	83

# **Planning/Evaluation**

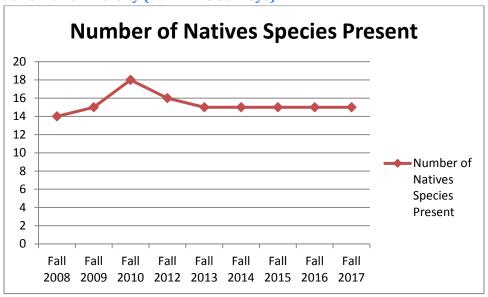
Surveys of the aquatic vegetation of the lake were conducted on June 1, 14, 21, July 26, August 2, 15, and September 18, 2017. Surveys of the lake were made frequently throughout the summer months for pre or post treatment evaluation, to collect water quality parameters, as well as to have additional survey data available for management purposes. Vegetation surveys determine the locations of target and non-target plant species. The results of the surveys are used to determine the most appropriate management strategy. The vegetation surveys also document the success of the prescribed management program. An AVAS survey is the State of Michigan's method for conducting a complete aquatic vegetation survey. The Aquatic Vegetation Assessment Site (AVAS) survey divides the parts of the lake capable of growing plants (littoral zone) into subareas and records the cover of each aquatic plant found in each "site". This method of surveying takes into account not only the types of plant species present in the lake but also the densities of those species. AVAS surveys are also an excellent way to track plant species trends over time. A goal of invasive plant management is to have native plants increase while exotic plants decrease over time. The success of this goal can be illustrated using the AVAS data collected over several years. Since different native plants grow at varying times throughout the season, it is important to evaluate the lake multiple times to account for all species in the lake. The first evaluation is conducted in the spring and is used to determine areas that will require treatment or management. The second survey is conducted in late summer or fall and is used to determine management success.

Table 3: Plant Species Found in Portage Lake -2017 \* Based from boat survey, not as precise as a walking shoreline survey

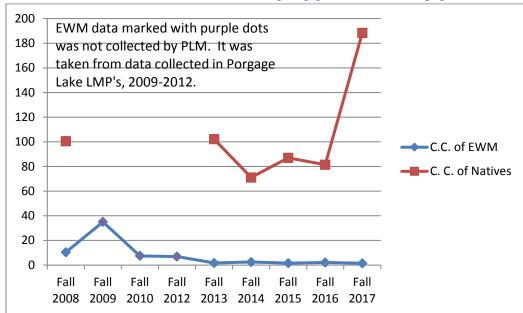
AVAS Code	Common Name	mon Name Scientific Name		% Cumulative Cove September 2017		
	Submerged- Exotic					
1	Eurasian watermilfoil	Myriophyllum spicatum	3.80	1.52		
2	Curlyleaf pondweed	Potomageton crispus	3.39	0.00		
	Submerged- Native					
3	Muskgrass	Chara	35.65	37.72		
4	Thinleaf pondweed	Potomageton spp.	10.33	6.74		
5	Flatstem pondweed	Potomageton zosteriformis	0.87	2.61		
6	Robbins pondweed	Potomageton robbinsil	0.00	0.00		
7	Variable leaf pondweed	Potomageton gramineus	0.22	1.41		

	Total		174.63	257.54
44	Common reed	Phragmites	1.76	3.48*
43	Purple loosestrife	Lythrum salicaria	0.65	0.54
	Emergent - Exotic			
42	Swamp loosestrife	Dianthera americana	0.00	0.00
40	Bulrush	Scirpus spp.	29.46	32.95
39	Cattail	Typha spp.	20.76	32.18*
30	Water lily	Nymphaea odorata	0.43	0.00
	Emergent- Native			
28	Nitella	Nitella flexilis	0.00	0.00
27	Sago pondweed	Potomageton pectinatus	7.17	13.15
25	Naiad	Najas flexilis	3.37	19.25
22	Bladderwort	Utricularia valgaris	1.96	1.85
21	Elodea	Elodea Canadensis	6.85	4.67
20	Coontail	Ceratophyllum demersum	2.39	6.21
17	Northern milfoil	Myriophyllum sibiricum	0.00	1.20
15	Wild Celery	Vallisneria Americana	14.89	38.70
11	Largeleaf pondweed	Potomageton amplifolius	1.85	5.76
10	Illinois pondweed	Potomageton illinoensis	16.10	25.11
9	Richardsons pondweed	Potomageton richardsonii	11.52	19.35
8	White stem pondweed	Potomageton praelongus	1.21	3.15

**Graph 1: Native Plant Diversity (Fall AVAS Surveys)** 



This graph shows the diversity of native plants found in Portage Lake. Portage Lake has excellent native plant diversity and this has been maintained throughout managing the exotic species.



Graph 2: EWM & Native Plant Cumulative Cover (C.C.) (Fall AVAS Surveys)

This graph shows the cumulative coverage of EWM from 2008-2017 as well as the overall cumulative coverage of all native plants in Portage Lake. The overall decline in the presence of EWM from the start of the management program shows the success of the treatments and that the population is currently being maintained at very low levels. The native plant population will naturally vary from year to year based on weather, water depth and many other factors; but has been maintained during the treatment of EWM. Please note that the EWM data marked with purple dots was data collected from another firm and not by PLM. This information was taken from the Portage Lake Forever website and used with permission of the board.

# **Genetic Testing/Sampling on Portage Lake**

Previous sampling on Portage Lake as shown that hybrid milfoil is present. Portage Lake was sampled twice in 2015 as part of PLM's participation in a grant program to sample and test milfoil plants for genetic analysis and herbicide sensitivity. More information on the grant study below:

From PLM's Spring 2015 PLM News Newsletter: "Recently the State of Michigan developed a "Michigan Invasive Species Grant Program" to be implemented in 2015 and is intended to be ongoing. Over 4 million dollars has been awarded to 20 different initiatives related to invasive plant management. Although all of these projects have relevant goals, PLM Lake & Land Management Corp



(PLM) understands the urgencies to utilize science to ensure balance of our aquatic ecosystems. Under the direction of Dr. Casey Huckins, Michigan Technological University (MTU), in partnership with Many Waters LLC., SePRO Corporation and PLM Lake & Land Management Corp; a grant application was submitted and approved for \$332,000. Although not every waterbody that we currently manage is directly involved in this project, PLM cliental representation is found throughout Michigan. To oversimplify; milfoil plant samples will be collected from over 15 different water bodies during the 2015

season. Samples will be sent to MTU for genetic analysis (providing specific hybrid genotypes of milfoil). Samples will also be sent to SePRO Corporation to simultaneously determine herbicide sensitivity of each hybrid type. Ultimately, we plan to verify the



specific genotype of milfoil and determine how we can effectively control it. If we do not determine an effective prescription for the control of certain genotypes of milfoil, we could potentially end up with a tolerance issue or select for herbicide resistant hybrid strains. For nearly a decade PLM has proactively implemented management protocols that rotate different types of herbicides at higher rates to reduce tolerance and resistance

potential, stay tuned. There are several other "multifaceted" objectives within this proposal."

Year 3 of this study is complete and no official updates are available currently.

#### Current Conditions in the Lake

# **Aquatic Vegetation**

Over the years, the presence of Eurasian watermilfoil and curly leaf pondweed undoubtedly reduced native plant diversity in the lake. Curlyleaf pondweed, although aggressive, naturally dies out mid-season and the increase in native plants after that die off is evident when looking at the early and late season surveys. Native plants currently have a good diversity and density in the lake.

Native plant diversity will continue to be promoted in the lake. The native plant species in Portage Lake benefit the lake, performing such functions as stabilizing sediments and providing habitat for fish and other aquatic organisms. In general, native species cause few problems, compared with those caused by exotic plants. Plant diversity is key to maintaining and improving the overall ecological balance of Portage Lake.

All of the plants listed in Table 3 are native North American species except Eurasian watermilfoil, Curlyleaf pondweed, Purple Loosestrife and Phragmites. These plants are non-indigenous aquatic nuisance species, i.e., plants from other places. These exotic plants cause considerably more problems than most native species. Eurasian watermilfoil can attain nuisance levels of growth at almost any time of year, whereas curly leaf pondweed completes its lifecycle and drops out of the water column by approximately the Fourth of July.

The native plant species benefit the lake, performing such functions as stabilizing sediments and providing habitat for fish and aquatic organisms. In general, native species cause few problems, compared with those caused by exotic plants. Three species commonly found in Portage Lake:



#### **Water Quality Monitoring**

Water quality monitoring is a critical part of lake management. Water quality monitoring provides an ongoing record of conditions in a waterbody. Changes in water quality can indicate threats from sources such as failed or inadequate septic systems, agricultural and lawn runoff, burgeoning development and erosion from construction site. Prompt identification of threats to water quality makes it possible to remedy them before irreversible harm has been done. Riparian's enjoyment of the water resource and the value of their property depend on maintaining water quality. The following tables break down the parameters tested in the different locations in Portage Lake including the Deep Hole Basins (Basin 1 and Basin 2), Shoreline Sites (3A, 3B, 3D), Tributaries (Glen Creek, McCormick Creek, Onekama Creek, Schimke Creek, Dunham Creek, Stream #9, Hansen Creek) and Storm Drains (#2, #5, #6, #7).

The graphs and tables below contain historical water quality data on Portage Lake that has been collected from numerous parties other than PLM. All information was made available to PLM via the Invasive Species Committee, on behalf of the Portage Lake Watershed Forever and Onekama Township and used with permission.



**Map 4: Portage Lake Water Quality Testing Locations** 

Table 4: Tributary Water Quality Portage Lake -2017 -cloudy/sprinkles/60
7 Temp D O Conduct- TDS pH TP ORP Turb TKN N

5/1/2017	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	Flow (Ft/sec)
Glenn	8.7	10.68	301.2	284.75	8.33	7	201	30.3	1.34	1310	27	1
McCormick	9	9.5	308.5	289	8.12	15	134	7.4	0.72	680	32	1.3
Onekama	9	10.86	286.8	268.95	8.44	8	183	8.6	1.21	1150	52	0.8
Schimke	9.7	10.65	270	248.21	8.33	7	146.3	6.5	1.8	1800	54	1.7
Dunham	9.1	10.95	271.3	253.2	8.49	8	159.5	0.5	0.94	911	21	1.4
Hansen	9.1	10.05	313.5	293.27	8.19	10	194.1	60.6	0.78	710	71	0.4
Stream #9	10.3	10.43	216.2	195.23	8.26	10	174.9	11.5	0.723	660	35	0.5
9/18/2017	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	Flow (Ft/sec)
9/18/2017 Glenn	•		ivity								onia	
	(C)	(mg/L)	ivity (uS/cm)	(ug/L)	(S.U.)	(ug/L)	(mV)	(NTU)		(ug/L)	onia	(Ft/sec)
Glenn	(C) 11	(mg/L) 10.55	ivity (uS/cm) 319.6	(ug/L) 283.3	(S.U.) 8.47	(ug/L) 35	(mV) 212	(NTU) 5.2	(mg/L)	(ug/L) 1250	onia (ug/L)	(Ft/sec) 0.2
Glenn McCormick	(C) 11 13.2	(mg/L) 10.55 9.85	ivity (uS/cm) 319.6 316.4	(ug/L) 283.3 262.2	(S.U.) 8.47 8.4	(ug/L) 35 22	(mV) 212 172	(NTU) 5.2 6.2	(mg/L)	(ug/L) 1250 1101	onia (ug/L) - -	0.2 0.4
Glenn McCormick Onekama	(C) 11 13.2 11.4	(mg/L) 10.55 9.85 10.28	ivity (uS/cm) 319.6 316.4 319.5	283.3 262.2 279.6	(S.U.) 8.47 8.4 8.57	(ug/L) 35 22 15	(mV) 212 172 202	5.2 6.2 4.2	(mg/L)	(ug/L)  1250  1101  230	onia (ug/L) - -	0.2 0.4 0.1
Glenn McCormick Onekama Schimke	(C) 11 13.2 11.4 12.1	(mg/L)  10.55  9.85  10.28  10.34	ivity (uS/cm) 319.6 316.4 319.5 355.8	(ug/L) 283.3 262.2 279.6 289.8	8.47 8.4 8.57 8.51	(ug/L)  35  22  15  15	(mV) 212 172 202 261	5.2 6.2 4.2 5.1	(mg/L)	1250 1101 230 1140	onia (ug/L) - - -	0.2 0.4 0.1 0.2

Table 5: Deep Hole Basin 1 Portage Lake -2017

(Secchi Disc: June 18', August 15', Sept.11')

Basin 1 June 1 2017	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	ALK (mg/L)	Chlor. A (ug/L)
S.	15.4	10.34	253	201	8.63	8	166	3.9	0.45	230	37	112	0.671
10'	15.3	10.33	253	201	8.6	-	176	0.6	-	-	-	-	-
20'	15.2	10.23	252	202	8.57	-	183	0.5	-	-	-	-	-
30'	12.2	11.17	230	198	8.51	6	189	0.4	0.307	280	20	118	0.94
40'	11.4	11.04	225	198	8.44	-	194	0.4	-	-	-	-	-
50'	11	10.52	224	199	8.37	-	199	0.5	-	-	-	-	-
60'	10.8	9.79	224	200	8.28	7	170	0.7	0.311	270	33	111	0.66
Basin1 Aug2 2017	Temp (C)	D.O. (mg/L)	Conduct- ivity	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia	ALK (mg/L)	Chlor. A
S.			(uS/cm)								(ug/L)	, ,	(ug/L)
	24	9.13	(uS/cm) 294	195	8.79	11	155	1.3	0.177	230	(ug/L) 47	103	(ug/L) 0.705
10'	24 23.7	9.13 9.3	• •	195 195	8.79 8.76	11	155 159	1.3	0.177	230		_	
			294									_	
10'	23.7	9.3	294 293	195	8.76		159	1.1	-	-		_	
10' 20'	23.7 18.6	9.3 9.9	294 293 262	195 195	8.76 8.53	-	159 169	1.1	-	-	47 - -	103	0.705
10' 20' 30'	23.7 18.6 14.3	9.3 9.9 8.9	294 293 262 243	195 195 198	8.76 8.53 8.18	-	159 169 179	1.1 0.7 0.6	- - .295	- - 230	47 - -	103	0.705

Basin1 Sep18 2017	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	ALK (mg/L)	Chlor. A (ug/L)
S.	20	9.89	270	194	8.97	11	148	1.8	0.102	230	45	114	1.05
10'	20	9.88	269	194	8.96	-	143	.09	_	_	_	_	_
20'	19.5	9.43	258	194	8.73	-	141	0.7	_	_	_	_	-
30'	13.1	7.58	235	201	8.13	10	140	0.9	0.134	230	22	119	0.488
40'	12	7.41	229	198	8.2	-	137	1.0	_	_	_	_	-
50'	11.5	7.3	227	199	8.14	-	131	1.2	_	_	_	_	_
60'	11.1	7.02	223.1	199	8.17	16	130	1.3	0.42	230	30	113	0.534

# Table 6: Deep Hole Basin 2 Portage Lake -2017 (Secchi Disc: June 20', August 16', Sept. 11')

Basin 2 June 1 2017	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	ALK (mg/L)	Chlor. A (ug/L)
S.	16.1	10.25	259	202	8.7	7	183	0.8	0.78	670	110	119	0.595
10'	16.1	10.26	259	203	8.71	-	186	0.6	_	_	_	_	-
20'	15.9	10.27	258	203	8.73	-	188	0.7	-	-	-	_	-
30'	15.8	10.22	257	203	8.69	7	193	0.7	0.19	230	25	117	1.04
40'	15.4	10.1	254	202	8.65	-	195	0.8	_	-	_	_	-
50'	14.2	9.96	247	202	8.54	-	201	0.6	_	_	_	_	-
60'	13.5	8.93	245	203	8.14	8	172	0.7	0.309	270	37	118	0.58
Basin2 Aug2 2017	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	ALK (mg/L)	Chlor. A (ug/L)
S.	24.6	9.17	299	195	8.63	11	186	1.5	0.124	230	18	109	0.417
10'	24.2	9.2	296	198	8.61	-	191	1.1	_	_	_	_	_
20'	20.2	8.21	277	199	8.25	-	204	0.9	_	_	_	_	-
30'	18.3	7.08	269	200	7.97	7	214	0.7	0.278	230	69	108	0.833
40'	17.3	2.45	275	209	7.51	-	226	1	_	-	_	_	_
50'	16.5	0.56	275	215	7.37	-	168	1.2	-	_	_	_	_
60'	16.3	0.5	276	216	7.36	13	160	1.2	0.443	230	368	122	0.224
Basin2 Sep18 2017	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	ALK (mg/L)	Chlor. A (ug/L)
S.	20.7	9.85	275	195	9	18	151	2.3	0.118	230	30	102	0.997
10'	20.2	9.92	227	194	9	-	146	1	_	-	-	-	-
20'	19.5	9.45	266	196	8.86	-	144	0.8	_	-	_	_	_
30'	18.4	8.29	264	196	8.72	16	144	0.8	0.123	230	27	101	0.552
40'	17.6	6.62	263	199	8.41	-	141	1	-	-	-	_	-
50'	17.31	5.15	246	201	8.2	-	133	1.1	_	_	_	_	_
60'	13.3	5.35	265	211	8.25	28	131	1.1	0.219	230	95	103	0.511

**Table 7: Shoreline Sampling Portage Lake -2017** 

Jun4 Secchi	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	ALK (mg/L)	Chlor. A (ug/L)
A 10'	16.9	9.93	269	207	8.65	7	166	1.1	0.257	230	39	121	0.641
B 6'	17.4	10.38	277	210	8.69	21	172	1.5	0.252	230	27	124	0.298
D 3'	17.3	10.02	289	221	8.59	8	165	2.1	0.393	360	29	138	0.336
Aug 1 Secchi	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	ALK (mg/L)	Chlor. A (ug/L)
A 12′	24.9	9.56	300	195	8.88	13	155	1.2	0.095	230	42	111	0.219
B 6'	25	10.15	300	195	8.88	12	151	1.3	0.147	230	24	108	0.0
D 3'	25.2	9.81	292	189	8.86	12	152	1.3	0.139	230	24	103	0.23
Sep18 Secchi	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	ALK (mg/L)	Chlor. A (ug/L)
A 8'	21.5	10.64	289	200	9.02	13	179	0.9	0.139	230	17	87	0.267
B 7′	21.6	10.07	289	201	8.86	11	185	0.8	0.124	230	31	83	0.828
D 4'	21.7	9.81	299	207	8.75	11	192	0.9	0.081	230	1	86	0.30

Table 8: Additional shoreline sampling sites Portage Lake - 2017

June1 Secchi	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	ALK (mg/L)	Chlor. A (ug/L)
1 2'	21.7	8.57	292	203	8.37	7	146	1.2	-	230	-	-	-
2 2'	22	9.28	293	203	8.41	10	139	0.9	-	230	-	-	-
3 2'	21.5	8.82	291	202	8.3	12	140	0.8	-	230	-	-	*
4 2'	22.4	8.65	298	204	8.27	15	142	0.8	-	230	-	-	-
Sep18 Secchi	Temp (C)	D.O. (mg/L)	Conduct- ivity (uS/cm)	TDS (ug/L)	pH (S.U.)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Amm- onia (ug/L)	ALK (mg/L)	Chlor. A (ug/L)
•			ivity								onia		Α
Secchi	(C)	(mg/L)	ivity (uS/cm)	(ug/L)	(S.U.)	(ug/L)	(mV)	(NTU)	(mg/L)	(ug/L)	onia (ug/L)	(mg/L)	Α
Secchi 1 2'	(C) 20.9	(mg/L) 10.08	ivity (uS/cm) 280	(ug/L) 197	(S.U.) 8.7	(ug/L) 16	(mV) 193	(NTU) 0.8	(mg/L)	(ug/L) 230	onia (ug/L) 12	(mg/L) 84	Α

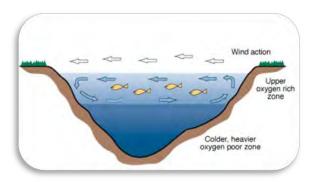
Table 9: Storm Drain Sampling Portage Lake - May 1, 2017

	Temp (C)	D.O. (mg/L)	Cond. (uS/cm)	TDS (mg/L)	pH (S.U)	TP (ug/L)	ORP (mV)	Turb. (NTU)	TKN (mg/L)	Nitrate (ug/L)	Flow (Ft/sec)	Weather post 2''rain
#2Zosel Park	9.7	7.43	327	300	7.86	12	69	0.5	0.922	830	0.6	Clear
#5 Fourth St	10.3	9.12	460	415	7.98	22	87	1.3	0.854	640	0.2	Clear
#6 Third St	10	10.19	303	278	7.99	23	77	2.8	0.587	440	0.2	brown
#7 First St.	10	5.65	366	335	7.49	33	24	12.2	1.157	380	0.1	clear

### **Temperature and Dissolved Oxygen Profiles**

Depth profiles of temperature and dissolved oxygen indicate that on June 1 the lake was already stratified. The surface levels were above saturation, 10.34 mg/L at Basin 1 and 10.25 mg/L at Basin 2 with shoreline ranging from 9.93 to 10.64 mg/L. At this time, Portage Lake had adequate dissolved

oxygen all the way down to 60' in depth (9.79 mg/L in Basin 1 and 8.93 mg/L in Basin 2). On June 1 the lake was thermally stratified, with a thermocline at approximately 30' in Basin 1 and 50' in Basin 2 slightly deeper than during a similar time in June of 2015 and 2016. The epilimnion (i.e., water above the thermocline) was well oxygenated, with oxygen concentrations at adequate levels to support a healthy fishery. Conditions in the hypolimnion (i.e., water below the thermocline) were also oxygenated.



On May 1, four storm drains (table 9) and seven tributaries (table 4) were tested coming into Portage Lake. All sites were well oxygenated ranging from 7.43 to 10.19 mg/L, except one site Storm Drain #7 at First St, which was at 5.65 mg/L. The oxygen level here was low, which was a similar reading as in 2016 and 2015. In the last four years, this storm drain has had very little flow, which could be impacting the oxygen levels. In 2017, the flow was very low. Storm Drain #5 and #6 had very little flow as well. This sampling was performed post a 2''rain event. Despise the rain event, flow was low and all but one drain was clear.

In August, the lake was still strongly divided. An August sampling was added into the program in 2015 and has been sampled since. Basin 1 was stratified and was anoxic at the bottom of the lake (void of oxygen). The thermocline in Basin 1 was 20' (whereas in 2016 it was at 30' and 2015 it was at 40'). Although the thermocline was higher, the oxygen levels stayed more consistent and didn't start declining until 40' and at that point the oxygen levels started a quick drop from 6.4 mg/L to 0.27 mg/L; anoxic water. The 2017 sampling showed more oxygen than the previous few years, which had shown a decline in trending oxygen levels. 3.0 mg/L is generally considered anoxic. In Basin 2, the surface waters had oxygen levels at 9.17 mg/L (similar to 2016 and 2015) and a thermocline at 20' (slightly higher than 2016 as well). Unlike in 2016 when DO levels dropped higher in the water column, the August sampling found oxygen levels dropping at 30' from 7.08 mg/L, at 40' 2.45 mg/L and 0.5 mg/L at the bottom (similar bottom reading to 2016).

In September, the lake was still stratified during the sampling period. In years past, both mixing and no mixing has been found during the September sampling. The warmer September weather allowed a strong stratification to be found in 2017, much stronger than in the last few years. Basin 1 was stratified at 30' and unlike in 2016 and other previous years, it was NOT anoxic below the thermocline (void of oxygen). DO levels ranged from 9.89 mg/L at the surface to 7.02 mg/L at the bottom. This is an excellent sign for the lake since 3.0 mg/L is generally considered anoxic. In Basin 2, which in many years has already mixed, had a strong thermocline during the sampling. Further, the oxygen was saturated from top to bottom, 9.85 mg/L at the surface, 8.29 mg/L in the thermocline and 5.35 mg/L at the bottom.

Substantial oxygen demand leads to rapid deoxygenation of the hypolimnion upon thermal stratification in the spring and oxygen concentrations are frequently decreased in bottom waters during the summer. Depletion of oxygen beneath the thermocline during the summer is a common symptom of eutrophication, and often leads to elevated internal nutrient loading as the result of the release of phosphorus from hypolimnetic sediments. The 2017 sampling shows better oxygen levels present in the hypolimnion.

#### pН

pH describes the balance between acids and bases in the water. Neutral values of pH are desirable. Low pH values typically result either from the growth of bog vegetation (such as peat moss), acid precipitation ("acid rain"), or acid runoff (as in acid mine drainage). Excessive growth of certain plants and algae can raise pH values. A majority of Michigan lakes have pH values in the 7-9 range. Portage Lake pH was recorded in Basin 1 and Basin 2 in the June, August and September as well as in the tributaries and shoreline sites. The pH in June ranged 8.14-8.73, in August from 7.37-8.79 and in September from 8.13 -9.00. The shoreline sampling was similar to the deep hole basins as was the tributary and storm drain sampling. This data is consistent with 2016 data as well as previous samplings.

#### **Total Alkalinity**

Alkalinity, in addition to pH, measures the amount of dissolved bases and the balance of acids and bases in the water. Alkalinity specifically measures the concentration of carbonates and bicarbonates in the water. These compounds and other ions associated with them can make water "hard". High alkalinity lakes are hardwater lakes, while low alkalinity lakes are softwater lakes. Different kinds of plants, algae and other aquatic organisms live in hardwater versus softwater. Alkalinity is a basic characteristic of water and is neither inherently good nor bad. Total Alkalinity was measured in June, August and September in both Basin 1 and Basin 2. The average sampling between both basins in June was 115 mg/L with a range of 112-119 mg/L. The August samples were similar with an average of 112 mg/L with a range of 103-123 mg/L. The September samples were similar with an average of 108 mg/L with a range of 101-119 mg/L. All samplings show the lake to be considered "soft" with readings under 150 mg/L, a typical threshold of a hardwater lake. Overall, the 2017 readings on the lake are slighter lower than previous readings, but overall show consistent softwater data for Portage Lake.

#### **Conductivity and Total Dissolved Solids**

Conductivity and Total Dissolved Solids (TDS) measure the total amount of material dissolved in the water. Higher values indicate potentially rich, more productive water, whereas lower values indicate potentially clean, less productive water. (If nutrient pollution is occurring, the total phosphorus concentration is a much better indicator of potential productivity.) The combined readings of TDS on Portage Lake ranged from June readings averaging 201 ug/L, August averages of 203 ug/L to September readings averaging 197 ug/L. (Shoreline samplings were very similar to deep basins). The tributary sampling was slightly higher, averaging 261 ug/L in May and 251 ug/L in September. Overall, these averages classify the overall TDS of Portage Lake as Low Dissolved material. The conductivity readings on Portage Lake are slightly higher than the TDS readings with the basin average of 245 uS/cm in May, 271 uS/cm in August and 237 uS/cm in September. (uS/cm=microsiemens per centimeter). Higher levels can likely be due to runoff, which is also supported by the slightly higher conductivity readings from the Tributaries (May average Conductivity reading is 280 uS/cm while September average is 291 uS/cm). Tributary readings are slightly higher than 2016, but generally speaking quite consistent.

#### Oxidative Reduction Potential (ORP)

The oxidative reduction potential of a lake measures the ability of the water to serve as potential oxidizers and indicates the degree of reductants present within the water (the ability to gain or lose electrons). The reduction potential measurement has proven useful as an analytical tool in monitoring changes in a system rather than determining their absolute value. Like pH, the redox potential represents an intensity factor. It does not characterize the capacity of the system for oxidation or reduction; in much the same way that pH does not characterize the buffering capacity. Generally speaking, higher ORP values, the healthier the lake. As a lake stratifies and oxygen levels decrease towards the bottom of the lake, ORP values will decrease even in a healthy lake due to the lack of oxygen. This is because there are many bacteria working in the sediments to decompose the material and they use up the available oxygen. ORP is measured in addition to pH and dissolved oxygen as it can provide additional information of the water quality and degree of pollution, if present. High ORP values indicate high levels of oxygen in the water and that bacteria that decompose the dead matter can work more effectively. The deep basins ranged from 166-201 mV in the spring sampling to 155-226 mV in August to 130-151 mV in September, indicating oxidized conditions. Tributaries and shoreline samples had similar results and these are similar readings to past samplings.

#### **Turbidity**

Turbidity is a measure of the clarity of the water, specifically from the presence of suspended particles in the water. Turbidity will typically increase as the suspended particles in the water increase, lowering clarity of the water. Turbidity may be caused by a variety of factors from the bottom sediments, erosion, algae production, and runoff and possibly from fish species such as carp. Suspended particles can capture heat from the sun raising water temperature as well (often witnessed in shallow waters). Turbidity readings on Portage Lake averaged 0.85 NTU's in June to 1.07 NTU's in August to 1.13 NTU's in September with similar readings throughout the water column. Shoreline sampling averaged was 1.5 NTU's in June, 1.2 NTU's in August and 0.86 NTU's in September. The tributaries average was overall higher, which would be expected in a shallow, flowing system (17.9 NTU's average in May and 9.4 NTU's in September). The tributary readings are much higher than previous years. During the May sampling, Glenn, Hansen and Stream #9 were outliers compared to historical data (30, 60,11 NTU's) During the September sampling, Hansen Creek and Stream #9 had a high reading of 34 and 10 NTU's. In 2016, Stream #9 was the only elevated reading. Rain can affect turbidity and a rain event had occurred prior to the spring sampling. Generally, more mixing occurs in shallow water, closer to the substrate. The World Health Organization (WHO) requires drinking water be less than 5 NTU's, but recreational water can be significantly higher. Overall, the turbidity readings on Portage Lake are within safe drinking water standards and overall show that clarity should be very good on the lake.

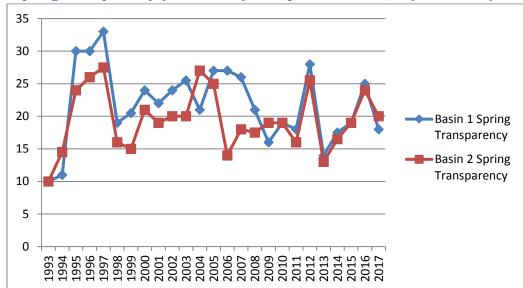
#### Secchi Disk Depth

The Secchi disk depth is another measure of water clarity, determined by measuring the depth to which

a black and white disk can be seen from the surface. (Larger numbers represent greater water clarity.) In June, Basin 1 was 18 feet while Basin 2 was 20' (slightly lower than 2016 recordings). Basin 2 is likely more affected by the fetch of the lake, therefore would likely have a lower Secchi disk reading, which has been seen in the past, but not during the 2017 samplings. Clarity declined with the Secchi disk depth of 15' in August in Basin 1 and 16' in Basin 2 (slightly higher than 2016) and was at 11' in Basin 1 and Basin 2 was at 11' in September (same as 2016). Water clarity can fluctuate from week to week depending on several environmental factors such as rain fall & algal production. These clarity readings show that sunlight will be available for plant and algae

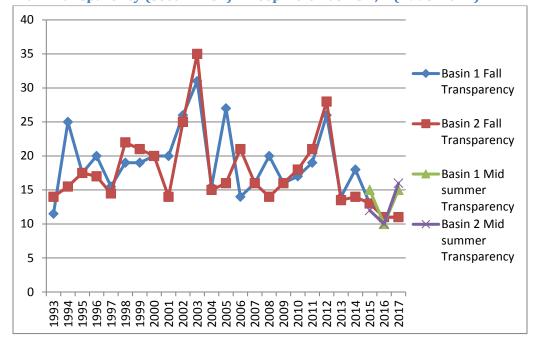


throughout the good portions of the lake. The shoreline sampling sites had very good clarity, with all readings reaching the bottom of the lake in both the June and September samplings.



Graph 3: Spring Transparency (Secchi Disk) - Deep Hole Basins 1, 2 (1993-2017)





#### **Total Phosphorus**

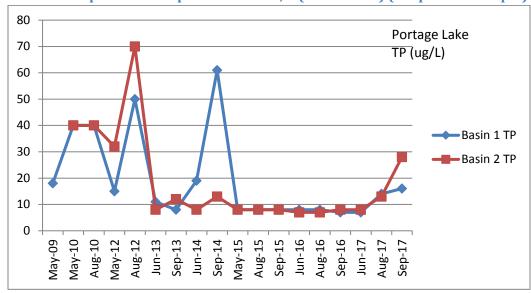
Total phosphorus measures the total amount of phosphorus in the water. Phosphorus is an important plant nutrient (i.e., fertilizer) and the nutrient most likely to limit algal growth. Phosphorus levels are not only related to internal loading of nutrients but also from external sources. Elevated phosphorus inputs to lakes caused by human activities are a major cause of cultural eutrophication. Total phosphorus concentrations in June in Basin 1 were 8 µg/L at the lake surface, and 6 µg/L at thermocline depth and 7 μg/L in the bottom water. In Basin 2, 7 μg/L at the lake surface, and 7 μg/L at thermocline depth and 8 μg/L in the bottom water. The June shoreline readings from sites 3A was 7 μg/L, 3B was 21 μg/L and 3D 8 μg/L. The tributary TP readings in May ranged from 7-15 μg/L. Storm Drain TP May readings were from 12-33 μg/L. Readings above 10μg/L are considered slightly enriched while readings over 30 μg/L are considered enriched. In the past, higher TP readings have been found coming from the tributaries and storm drains. Overall, the spring samplings on the lake have stayed similar to 2016 and 2015 reading,

showing a slight trend down. The tributaries and storm drains had slight increases at some locations, but no values of a large concern.

August Total Phosphorus concentrations were: Basin 1: 11 μg/L at the surface, 8 μg/L in the thermocline and 14 μg/L at bottom while Basin 2: 11 μg/L at the surface, 7 μg/L in the thermocline and 13 μg/L at bottom. Slight increases from the June testing but all readings still well below levels of concern.

September Total Phosphorus concentrations were: Basin 1 11 μg/L at the surface, 10 μg/L in the thermocline and 16 µg/L at bottom while Basin 2 18 µg/L at the surface, 16 µg/L in the thermocline and 28 µg/L at bottom. All of these results are slightly higher than 2016. 2015 and 2014 data showed lower TP results with a slight increase in 2017, even though more oxygen was present during much of the sampling periods. Note: The levels are still under the 30 μg/L level, which we hope to avoid. The shoreline readings from sites 3A (13  $\mu$ g/L), 3B (11  $\mu$ g/L) and 3D (11 $\mu$ g/L) while the tributaries overall ranged from 13 μg/L to 35 μg/L. In years past, Stream #9 was the highest of the readings, in 2016 McCormick showed the higher reading, and in 2017, Glenn Creek had the highest reading at 35 μg/L. The shoreline and tributary September samplings showed higher levels of TP, with numerous results considered slightly elevated-to-elevated. Additional tributary upstream testing done in September did find slightly elevated phosphorus levels in Stream 9 (22 μg/L -34 μg/L). Stream #9 was the only location sampled based on high results found in 2016. The September readings show that overall, slightly higher phosphorus concentrations are found in the tributaries and that internal loading was not a large contributing factor to TP in 2017 (or in 2016). The 2017 data shows the TP has increased slightly in both Basins, but still well below historical data that has been elevated. Past data has shown that Basin 2 is routinely higher in concentrations than Basin 1, which is expected due to the fetch and potential lack of oxygen of Portage Lake.

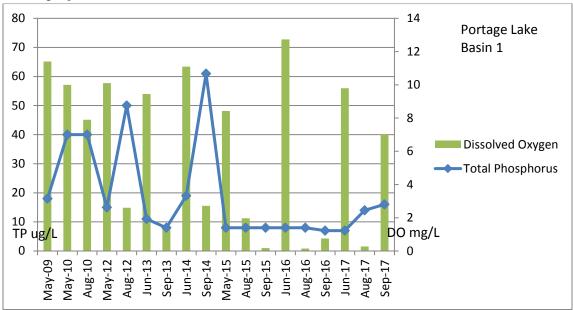
See below graphs of TP concentrations from 2017. Basin 1 and 2 are graphed using data previously collected on Portage Lake (via various sources, provided to PLM via the Portage Lake Watershed Forever website with permission from the committee).



Graph 5: Total Phosphorus - Deep Hole Basins 1, 2 (2009-2017) (deep water sample)

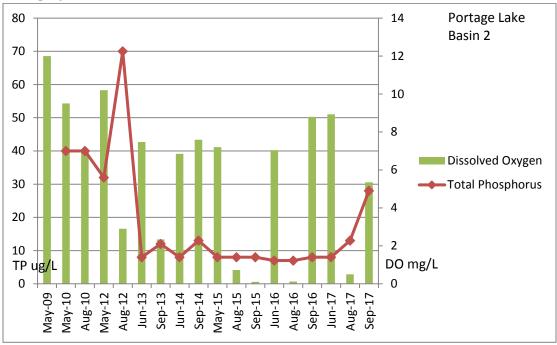
There have been a few spikes in TP over time, an overall decrease in 2013 and a large spike in Basin 1 in 2014, likely an outlier based on 2015 and 2016 results. Basin 2, which has been higher in TP, had the same results in 2015 and 2016, showing a DECLINING trend in overall TP in Portage Lake! The 2017 results, although show a trend up, are far below data collected from 2009-2012. Note: Basin 2 May 2009 sample is not graphed as the reading of 340 ug/L is an extreme outlier and not reflective of the overall lake results.

Graph 6: Total Phosphorus & Dissolved Oxygen - Deep Hole Basin 1, (2009-2017) (deep water sample)

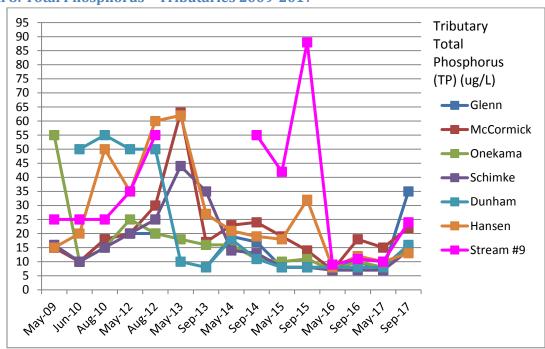


Internal loading can take place when dissolved oxygen levels decrease. 2015 and 2016 results show decreased DO levels, which again can cause internal loading, but no evidence of any increases in TP concentrations are found. In 2017, DO levels throughout the summer were better, but a slight increase in TP concentrations was found.

Graph 7: Total Phosphorus & Dissolved Oxygen - Deep Hole Basin 2, (2009-2017) (deep water sample)

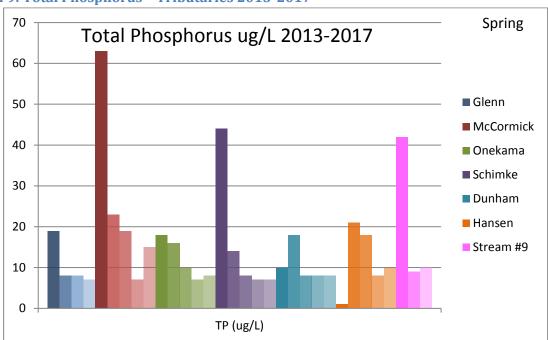


DO levels have decreased the last few years, and up until late summer 2017, TP levels had not increased. However, the August and September TP levels show an increase with both saturated and un-saturated water present. Increase TP levels with low oxygen can be an indicator of internal loading. Continued monitoring recommended.



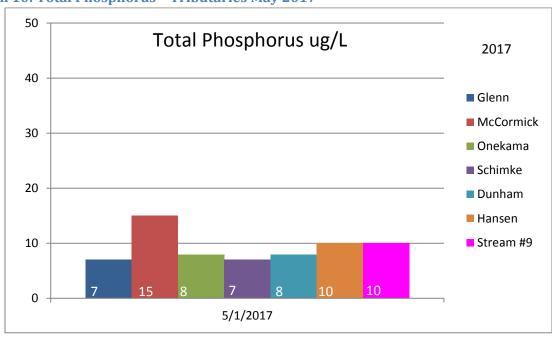
**Graph 8: Total Phosphorus - Tributaries 2009-2017** 

As the graph illustrates, there are fluctuations between the creeks over time. See below graphs to show the 2017 comparisons between the creeks. Glenn Creek May 2013 sample was removed from this graph as an extreme outlier, likely from a contaminated sample. Stream#9 was not sampled in 2013 but is currently showing the highest TP concentrations among the Tributaries.



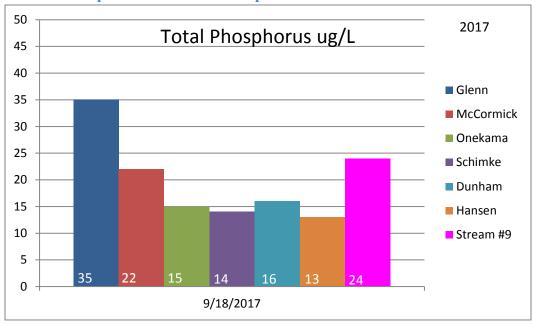
**Graph 9: Total Phosphorus - Tributaries 2013-2017** 

As the graph illustrates, there are fluctuations between the creeks over time. See below graphs to show the 2017 comparisons between the creeks. Glenn Creek May 2013 sample was removed from this graph as an extreme outlier, likely from a contaminated sample. Stream #9 was not sampled in 2013 but is currently showing the highest TP concentrations among the Tributaries.



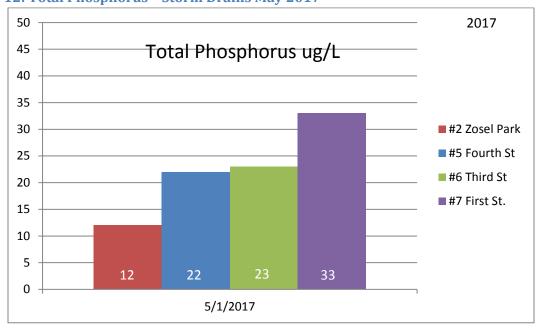
**Graph 10: Total Phosphorus - Tributaries May 2017** 

As the graph illustrates, very little fluctuation between the TP in the different creeks entering Portage Lake was found in 2017 (same for 2016). In years past, concentrations have ranged more and been more enriched.



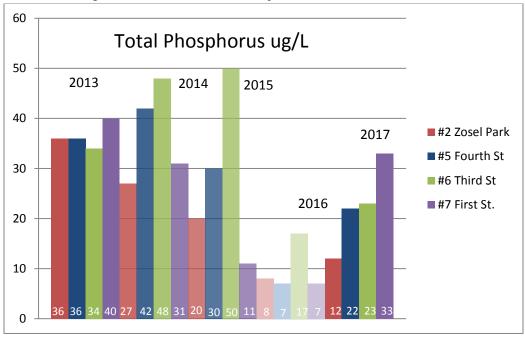
Graph 11: Total Phosphorus - Tributaries September 2017

As the graph illustrates, there is fluctuation between the TP in the different creeks entering Portage Lake and overall, the samples are more enriched than 2016.



**Graph 12: Total Phosphorus - Storm Drains May 2017** 

As the graph illustrates, there is a fluctuation between the TP in the different storm drains around Portage Lake and overall, the samples are higher than the last few years.

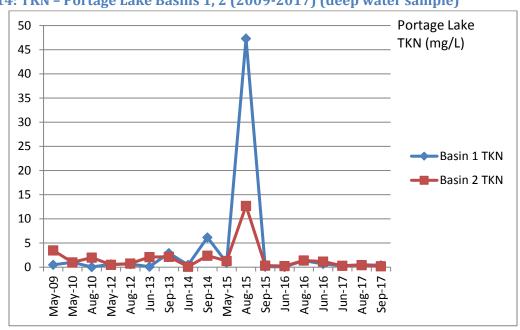


Graph 13: Total Phosphorus - Storm Drains May 2013 - 2017

As the graph illustrates, there is variance between the TP in the different storm drains entering Portage Lake yet all the TP concentrations are considered enriched. These sites are a key introduction point of Phosphorus into Portage Lake. In general, drains have decreased in 2015 and 2016, with increases in 2017. Although the levels are lower than historical data, increases were found in 2017.

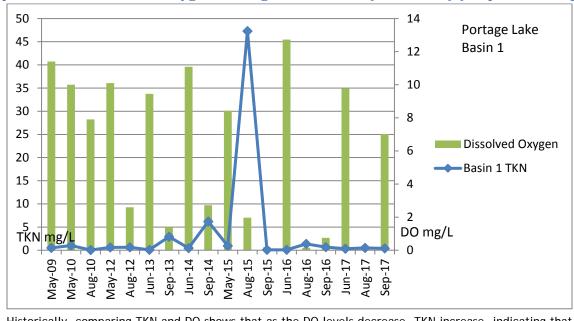
#### **Total Kjeldahl Nitrogen (TKN)**

TKN measures the total organic amount of nitrogen (nitrate and nitrite) and ammonia in the water. Nitrogen is the plant nutrient (i.e. fertilizer) most likely to control the amount of rooted plant growth in lakes and ponds. Most Midwestern lakes have more nitrogen and more rooted plant growth than is desirable, so lower values are generally considered better. The major sources of nitrogen in lakes are from agriculture (animal waste, fertilizer) and atmospheric emissions (fossil fuel). Lakes with a TKN value of 0.66 mg/L or less are typically classified as oligotrophic lakes (having fewer nutrients, less productivity). Lakes with TKN values above 1.88 mg/L may be classified as eutrophic (highly productive and nutrient rich). Nitrates do not accumulate very much in the bottom waters during the summer because when nitrate is void of oxygen it turns into ammonia. Therefore, ammonia testing is an excellent way to determine internal loading of nitrogen. The TKN readings on Portage Lake at Basins 1 and 2 in June ranged from 0.19 mg/L to 0.73 mg/L, in August from 0.124 mg/L to 0.495 mg/L and in September from 0.102 mg/L - 0.402 mg/L between both basins. These readings are slightly lower than in 2016. The tributaries and storm drains showed lower levels than past years as well. The tributaries samples ranged from 0.72 mg/L-1.8 mg/L in May while the Storm Drains ranged from 0.58 mg/L - 1.15 mg/L in May. TKN readings decreased in 2017 from levels that had increased in 2015 and continuing to test this parameter is recommended.



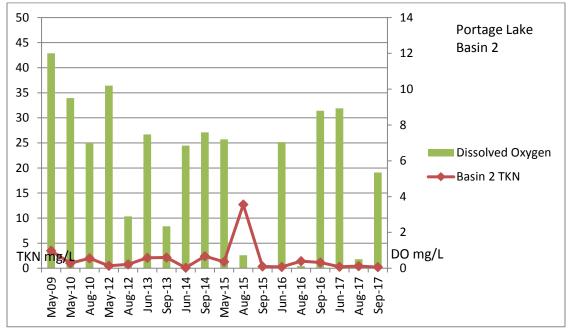
Graph 14: TKN - Portage Lake Basins 1, 2 (2009-2017) (deep water sample)

As the graph illustrates, the TKN concentrations on Portage Lake have fluctuated over the last few years, with a spike in August 2014 and a large spike in August 2015, but have returned to lower, less enriched levels in all of the 2016 and 2017 readings.



Graph 15: TKN & Dissolved Oxygen-Portage Lake Basin 1 (2009-2017) (deep water sample)

Historically, comparing TKN and DO shows that as the DO levels decrease, TKN increase, indicating that internal loading is likely taking place. However, in 2015, low DO levels correlate with low TKN levels in September, but not August. 2016 samplings show low TKN levels regardless of DO levels, indicating no internal loading taking place. 2017 data supports the 2016 conclusion.



Graph 16: TKN & Dissolved Oxygen-Portage Lake Basin, 2 (2009-2017) (deep water sample)

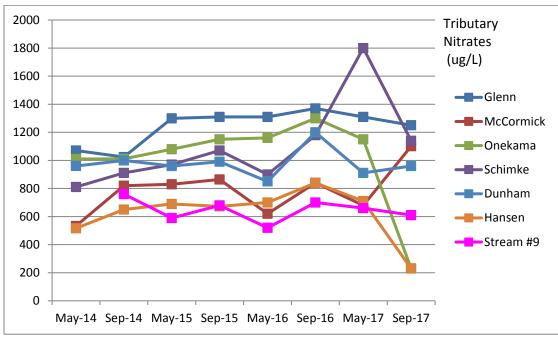
Similar 2015 results as in Basin 1. 2017 data supports the 2016 data conclusion that DO levels are not impacting TKN levels, leading to the conclusion of no internal loading.

#### **Nitrates**

Nitrates measure the total amount of in-organic nitrogen in the water. Again, nitrogen is an important plant nutrient (i.e., fertilizer) and the nutrient most likely to limit the growth of rooted plants. Most Midwestern lakes have more nitrogen and more rooted plant growth than is desirable, so lower values are generally considered better. Nitrate levels under 250 µg N/L are considered not enriched while readings between 250-750 μg N/L are slightly enriched, readings from 750-1250 μg N/L are enriched and

readings over 1250 µg N/L are highly enriched. The June concentrations of nitrates in Basin 1 and 2 ranged from 230 μg N/L to 670 μg N/L. The August and September concentrations of nitrates were all 230μg N/L in both basins throughout the water column expect for one reading at 291μg N/L in Basin 1 at the bottom in September. Both Basin results are down similar to 2016. Nitrates in the tributaries ranged from 660 μg N/ to 1800 μg N/L in the spring and from 230 μg N/ to 1250 μg N/L in September, which is similar to 2016. The Strom Drains had slightly lower readings than the tributaries (380 μg N/L to 830 μg N/L), classifying as slightly enriched with one drain enriched. Nitrates are typically higher in the spring when the water is colder because the bacteria needed to digest the nitrates are not as productive in cooler temperatures. Nitrates will often decrease over the spring and were slightly less in the lake by the end of the summer. Nitrate levels remained low throughout the rest of the season with an overall lake average of 235 μg N/L. Based on the higher levels of nitrates observed in inlets (Tributaries) in May and September, loading of the lake appears to be mainly from external sources. External sources for nitrate pollution are agricultural practices (manure, fertilizer), animal feedlots, urban runoff and municipal wastewater runoff. Based on the location of Portage Lake and the makeup of the surrounding watershed, nitrate enrichment is most likely coming from agricultural practices that have leached into the groundwater and animal feedlots. Nitrates did not accumulate very much in the bottom waters during the summer. The nitrates did not accumulate because when nitrate is void of oxygen it turns into ammonia. Therefore, ammonia testing is a better way to determine internal loading of nitrogen.

These samples show that the lake (at the time of sampling) may be Phosphorus limited. Phosphorus limited lakes tend to have a TN:TP >15. In 2017 the average TN was 265 ug/L in the basins and the TP 11 ug/L, giving a TN:TP of 24, indicating Phosphorus may be the limiting nutrient. This is common in most lakes in this geographical area.



**Graph 17: Nitrates-Portage Lake Tributaries** 

As the graph illustrates, the nitrate concentrations in the Portage Lake Tributaries range from slightly enriched to enriched to highly enriched in. It is recommended to continue testing.

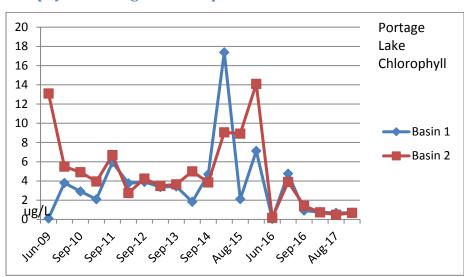
#### **Ammonia**

Ammonia is a form of nitrogen found in organic materials, sewage, and many fertilizers. It is the first form of nitrogen released when organic matter decays. Also, when ammonia degrades it consumes oxygen, which worsens already existing anaerobic conditions. However, ammonia can be used by most

aquatic plants and is therefore an important nutrient. When oxygen is present in a lake ecosystem, ammonia will convert to nitrates. Ammonia is toxic to fish at relatively low concentrations in pH-neutral or alkaline water. In fish, ammonia affects hatching and growth rates, and can cause changes in tissues of gills, the liver and the kidneys. Ammonia concentrations below 1000 ug/L are considered suitable for healthy fisheries. Ammonia concentrations usually do not become elevated until water is void of oxygen and the nitrates are converted. Therefore, concentrations of Ammonia did not become elevated until anaerobic conditions are present, typically mid-summer. The concentration of ammonia at the Basin 1 in June was 37 ug/L at the surface and 33 ug/L at the bottom while in Basin 2 it was 110 ug/L at the surface and 37 ug/L at the bottom. In August, the concentrations were 47 ug/L at the surface and 393 ug/L at the bottom in Basin 1 and 18 ug/L at the surface and 368 ug/L at the bottom in Basin 2. The September concentrations were 45 ug/L at the surface and 101 ug/L at the bottom in Basin 1 and 30 ug/L at the surface and 95 ug/L at the bottom in Basin 2. The hypolimnion (deep-water) concentrations observed in August had increased from June, but still under the threshold of concern. September readings are well within range for a healthy fishery as well. The shoreline areas ranged from 1-39 ug/L throughout the summer, all considered very low. As oxygen is not an issue here, this is expected. The tributaries had similar levels of ammonia as the lake throughout the season. Ammonia concentrations ranged from 21 ug/L to 71 ug/L in the tributaries. Storm drain levels were similar except for First Street #7, which had the highest reading of all areas at 766 ug/L. The other drains ranged from 79-201 ug/L. This drain had the lowest oxygen level at 5.6 mg/L, substantially lower than the rest of the sampling sites. As more ammonia was present here, it could have affected the oxygen levels as well.

#### Chlorophyll

Chlorophyll measures the amount of plankton (green plant) in the water. Some plankton or algal growth is essential to support the growth of other organisms (e.g., fish) in the lake, but human activities and natural eutrophication often lead to excessive algal growth; thus, lower concentrations of chlorophyll are usually considered desirable. Chlorophyll concentrations in Portage Lake Deep Basins in June ranged from 0.58 ug/L to 1.04 μg/L indicating low plankton populations. Shoreline samplings sites (3A, 3B, 3D) averaged 0.42 ug/L. Chlorophyll in the Deep Basins ranged from 0.224 ug/L - 0.876 ug/L. In September, Chlorophyll ranged from 0.488 ug/L to 1.05 ug/L. The shoreline sites averaged 0.465 ug/L. A higher level, in shallow, warmer waters is common as the warmer water can be a breeding ground for plankton. Overall, chlorophyll levels, which had increased in 2015, have lowered again in 2016 and 2017, similar to historical data for Portage Lake.



Graph 18: Chlorophyll a- Portage Lake Deep Basins

As the graph illustrates, overall Chlorophyll a sampling has declined over the last few years with some spikes, likely weather related.

#### **Algae and Zooplankton Composition**

Algal composition testing was performed at both deep Basins as well as the shoreline testing sites in June, August and September. The June testing showed the majority genera present included (presented as most abundant to least abundant); Bacillariophyta (diatoms): Cyclotella sp., Asterionella sp., Fragilaria sp.; Chlorophyta (green algae): Ankistrodesmus sp.. The August sampling found Cyanophyta (blue green algae), specifically *Microcystis sp.*, *Gloeotrichia sp.*, Gomphosphaeria sp., the most abundant species and genera of phytoplankton followed by Chlorophyta (green algae): Pediastrum sp., Chlorella sp., Zygnema., Ankistrodesmus sp.; Bacillariophyta (diatoms): Fragilaria sp., Cyclotella sp., Asterionella sp.. The September sampling found Cyanophyta (blue green algae), specifically Microcystis sp., Gloeotrichia sp., the most abundant species and genera of phytoplankton followed by Chlorophyta (green algae): Pediastrum sp., Chlorella sp.; Bacillariophyta (diatoms): Fragilaria sp., overall, concentrations were low. Some blue green algae, including Microcystis sp., can produce toxins. These toxins are normally released when the algae near the end of the life cycle and often occur for short phases during a growing season, often times towards the end of the season after the water temperatures and nutrient loading have reached a high. Further, blue green algae are not consumed by Zebra mussels, so if Zebra mussels are present in a lake ecosystem, it is likely to have lower green algae populations and higher blue green algae, as the Zebra Mussels will filter the green algae out of the water column and leave the blue green algae alone. The levels of blue green algae are not high enough to warrant a concern at this time. The blue green algae "scum" that forms on the lake surface when densities are extremely high should be avoided if that were to occur, but the densities in Portage Lake are not high enough to cause a bloom at this point. The zooplankton communities were also identified while looking at the phytoplankton and numerous species of zooplankton were documented including; Cladocera sp. (Daphnia)., Rotifer sp., Brachiopoda sp., and Copepods sp. Diverse and present phytoplankton is required to have a healthy zooplankton community as the base of the food chain.

#### Fecal Indicator Bacteria (E. Coli)

Fecal Indicator Bacteria (E. Coli) measurements count the number of live fecal indicator bacteria in the sample. These bacteria are considered reliable indicators of fecal contamination when they are found in a pond or lake; it is very likely that the water is being contaminated by animal feces. Contamination can potentially be derived from a number of sources, including failed septic systems, agriculture runoff, or waterfowl or wildlife droppings. E. Coli was tested in Portage Lake in August 2017. Four locations of concern were tested in the lake including the camp areas, Portage Point Inn, and a beach area on the SW end of the lake. All samples came back very low.

**Table 10: E. Coli Results in Portage Lake -2017** 

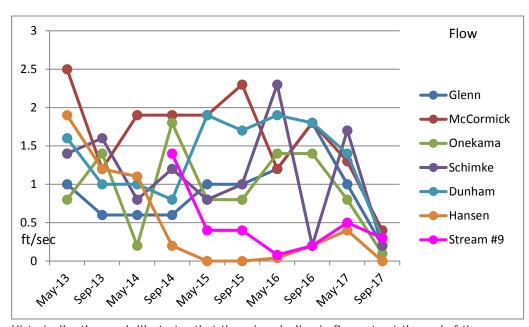
	E. Coli (CFU/100mL)	Total Coliforms (CFU/100mL)	Notes
Eden Camp	<1	2	Water meets bacteriological standards for safe swimming
Portage Pt. Inn	<1	435	Water meets bacteriological standards for safe swimming
Resident beach on SW side (active during sampling)	<1	411	Water meets bacteriological standards for safe swimming
Covenant Camp	1	687	Water meets bacteriological standards for safe swimming

Bacterial counts are expressed as the number of Colony Forming Units per 100 milliliters (CFU/100mL).

For full body contact recreation (including swimming) counts of E. coli should not exceed 130 (CFU/100mL) as a monthly geometric mean of at least five samples per the State of Michigan standard, or single samples should not exceed 298 (CFU/100mL) [235 CFU/100mL in a designated bathing beach area] per Federal (EPA) guidelines. Current recreational water quality standards do not rely on Total Coliform counts.

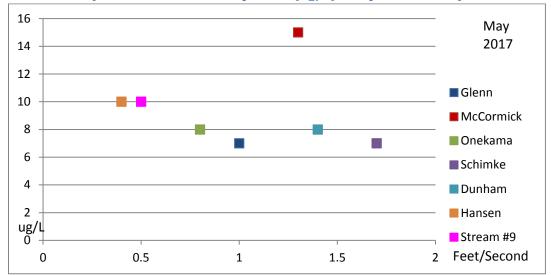
#### **Tributary Flow and Phosphorus**

Flow rate data was determined, using a digital flow meter, at the seven tributaries studied in May and in September 2017. Flow ranged from 0.4 feet/second - 1.7 feet/second in the May sampling and from 0.0 feet/second - 0.4 feet/second in September. Schimke was the fastest in May and McCormick, being the fastest in September, similar to past years. The rates of flow varied from each creek and the basic chemistry varied as well. Nutrients coming in from the creeks are a concern, as it is a transport from the watershed into Portage Lake. Total Phosphorus is graphed below along with flow to see how the flow and TP are connected.



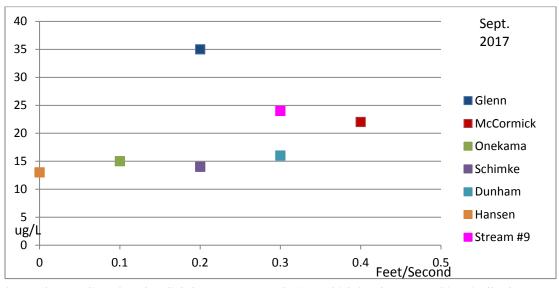
**Graph 19: Tributary Flow Rates - May and September 2013-2017** 

Historically, the graph illustrates that there is a decline in flow rate at the end of the summer versus the beginning of the summer. Typically, higher flows in spring will increase nutrient inputs in the spring and they decrease in the fall. This is standardly due to snow melt and spring rain. Generally speaking, the tributaries had less flow in September 2017 than previous years.



Graph 20: Tributary Flow Rates and Phosphorus (ug/L) comparisons -May 2017

In years past, the graph has illustrated a correlation between flow and TP. The greater the flow, the higher the Total Phosphorus. (Exceptions: Stream #9 is highly nutrient enriched.). This correlation has historically been strong. In 2016 and 2017, the TP concentrations are generally low across all tributaries, showing a smaller correlation.



Graph 21: Tributary Flow Rates and Phosphorus (ug/L) comparisons -September 2017

September readings found a slightly stronger correlation, which has been seen historically, between flow and TP.

#### Additional Tributary/Upstream testing

Tributary testing was expanded in 2016 to include testing four creeks upstream to determine if there were any point source locations determined or pinpointed. Determining any area of concern would allow future work to reduce nutrient loading into the lake be done. Using best management practices throughout the entire watershed, but especially on the creeks leading directly into the lake are essential.

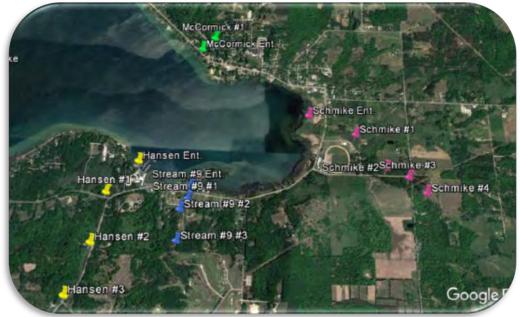
Determining if there is a location within the first few miles of the creek off of the lake that has elevated nutrient levels would allow future focus to be determined.

Based on historical data of nutrient levels from the tributaries, four creeks were selected to have additional testing done. Those creeks include: McCormick, Schimke, Hansen and Stream #9. During this test, each creek was also tested upstream at locations that were determined upon walking up the creek. Upon walking upstream, visual observations were made for any concerns including but not limited to drain tiles, erosions, buffers, invasive, flow issues, sources of nutrient inputs, etc. Based on observations the following locations were selected as potential sources of nutrient inputs: culverts, wetlands, location of golf course, farming field, houses, roads, etc.

Of the data collected, most locations came up somewhat enriched, with the largest concern being Stream #9. Because Stream #9 was the largest concern in 2016, it was selected for upstream testing in 2017.

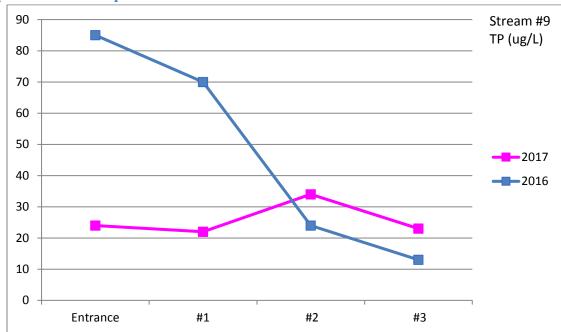
Of the four locations sampled on Stream #9, all samples came back lower than 2016 results, but are still considered enriched. Based on variances in the 2016 and 2017 data, it is likely indicative that it is highly impacted with flow and rain events.





**Table 11: Upstream Tributary Testing 2017** 

	9/18/2017	Total Phosphorus	Nitrates	Aikaiinity
Stream #9	Entrance	24	610	190
	#1	22	760	170
	#2	34	380	147
	#3	23	680	169

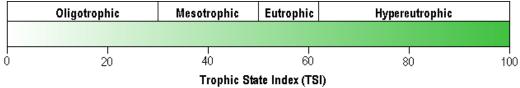


**Graph 22: Total Phosphorus Stream #9** 

Stream #9 had overall lower TP concentrations in 2017 than 2016.

#### **Evaluation of Trophic Status**

Carlson's Trophic State Index (TSI) is used to measure the trophic state of individual lakes. Lakes are ranked from 1 to 100 based on Secchi disc depth, Total phosphorus concentrations and/or Chlorophyll a levels. Based on that ranking, the TSI is determined. This chart gives the approximate classification for each category.



Portage Lake's June data yielded values between 28 and 36, in August between 24 and 38 and in September between 27 and 43 (Table 12). In general, these values rate Portage Lake as oligotrophic to mesotrophic. Characteristics associated with oligotrophic to mesooligotrophic lakes are low nutrient levels, clear water and low productivity. High dissolved oxygen levels typically occur and survival of cold water fish is possible. Mesotrophic lakes tend to have moderate nutrient levels, clear water and moderate productivity. Rooted plants are abundant and the lake can still support a cold water fishery. As the picture to the right shows, eutrophic lakes (not Portage Lake at this time, but given for comparison) have high nutrient levels, turbid water, algae blooms are likely and sometimes severe. Plants are abundant and dissolved oxygen is often depleted from bottom waters, restricting fish populations to warm water species.

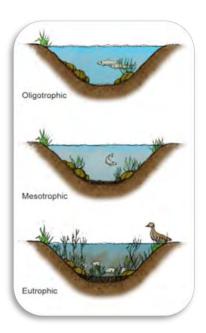


Table 12: 2017 Trophic State Index (TSI) Values

Site	Secchi Depth	Total Phosphorus	Chlorophyll a
Basin 1 - June	36	28	28
Basin 2 - June	34	28	28
Basin 1- Aug	38	34	27
Basin 2- Aug	37	33	24
Basin 1 - Sept	43	36	27
Basin 2 - Sept	43	43	27

### **2017 Water Quality Concerns/Recommendations**

Current water quality problems in Portage Lake can result from nutrient loading from the watershed and nutrient rich bottom sediments in the lake. Please note that the overall nutrient levels in Portage Lake are still relativity low compared to most Michigan waterbodies. Reductions in external nutrient loads may eventually reduce internally generated water quality problems, though improvements will require that dramatic reductions in external loading be sustained for long periods of time. Even if sufficient loading reductions are achieved, many years will be required before improvement is evident. In order to manage external nutrient inputs, it would be necessary to develop and implement a watershed management plan for the Portage Lake watershed. Watershed activities and public awareness using good management practices in the watershed will have long term positive improvements in the lake. This could be one cause of the decrease in nutrient levels in the lake.

# Management Recommendations for 2018

Management options are dependent on many factors, including but not limited to, species abundance (density), species richness, species location and many lake characteristics. Whenever an exotic species is found within an aquatic environment, action needs to be taken to prevent long term ecological damage as well as recreational and aesthetic loss that will take place.

#### **Submersed Aquatic Plants**

The 2018 aquatic plant management program should detect and treat any areas where Eurasian watermilfoil or hybrid watermilfoil are present in addition to any other invasive, exotic species.

Any areas of Eurasian watermilfoil should be promptly treated using herbicides. Treatments with the herbicides, Triclopyr and/or 2,4-D, in localized treatment areas to slow the spread of Eurasian watermilfoil, when found should be conducted. The herbicides Triclopyr and 2,4-D, control Eurasian watermilfoil with little or no impact on most native plant species. Since they are selective, systemic herbicides, they can actually kill Eurasian watermilfoil plants. Under ideal conditions, several consecutive annual applications of Renovate or 2,4-D can reduce Eurasian watermilfoil to a maintenance (low) abundance. For this strategy to succeed, it is necessary to treat all the Eurasian watermilfoil in the lake each time they are applied. Michigan regulation restricting 2,4-D use in the vicinity of drinking water wells may result in the inability to apply 2,4-D near the shoreline of the lake.

Triclopyr is a systemic herbicide with selectivity very similar to 2,4-D. Triclopyr is not subject to the well setback restrictions that currently affect 2,4-D. Therefore, triclopyr can be used to control Eurasian watermilfoil in near shore areas. A combination of both systemic herbicides in Portage Lake could greatly reduce the growing Eurasian watermilfoil problem.

Several contact herbicides, including diquat, can also provide short-term control of Eurasian watermilfoil. These herbicides kill only the shoots of the plant, and plants regrow relatively rapidly from their unaffected belowground parts.

Nuisance native plant management can also be incorporated into a lake management program with conventional herbicide treatments if needed. Native plant treatments are completed using only contact herbicides in beach areas. Contact herbicides will not target the root system of the plant.

# **Emergent Vegetation Management**

Purple loosestrife and Phragmites should continue to be addressed around the perimeter of the lake to prevent the further spread of these exotic species. The systemic herbicides, Glyphosate and Imazapyr, are effective at controlling Phragmites while Renovate 3 is effective in controlling Purple Loosestrife. Since they are systemic herbicide, the root system of the plant will be killed not just the foliage. Further, Purple Loosestrife should continue biological control measures as well. In addition, any other invasive terrestrial plants including but not limited to Japanese knotweed, honey suckle, garlic mustard and autumn olive should be targeted for control.

#### **Monitoring**

Aquatic vegetation and water quality should continue to be monitored to document the condition of the lake and to provide warning of any changes in the condition of the lake that need to be addressed by additional lake management activities.

# **Proposed Budget**

The following budget is proposed based on previous requirement on Portage Lake and the budget is limited to the management and treatment of Portage Lake. If additional costs are required in the maintenance of the SAD or from outside factors, they may not be included in this budget. Please also note that as additional data becomes available from the Grant Study and application rates increase, the budget may have to be adjusted long term to account for genetically changing plants.

**Table 13: Proposed 2018 Budget Portage Lake** 

Proposed/ Estimated Budget	2018
Emergent Control	5,000
EWM Control	54,500
Permit	1,500
Lake Management	15,000
Contingency Funds	7,600
Total	83,600

# The Recommended Management Schedule for 2018:

- A spring and fall vegetation survey (to evaluate conditions in the lake).
- Herbicide Treatment for exotics as required
- Pre and post treatment surveys as required, in addition to a mid-summer survey
- Extensive water quality monitoring throughout season
- Late summer/fall Phragmites treatment